

## Modulation of T cell response by *Phellinus linteus*

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***Phellinus linteus*, a species of mushroom, has been shown to contribute to health benefits, such as anti-inflammatory activity and immunomodulatory efficacy. The aim of this study was to analyze the most effective constituents of *P. linteus* fermented broths, polysaccharides, and to evaluate their immunoregulatory effects on T cells. Four fermented broths (PL1–4) and the dialyzate medium (MD) were prepared from *P. linteus* mycelia, and the polysaccharide contents of each were analyzed. The *P. linteus* samples were tested for biological activity in the regulation of T cell activation. In T cells, the production of mitogen-induced interleukin (IL)-2 and cell cycle progression were dose-responsively inhibited by PL3 and MD, primarily through cell-cycle arrest in S phase. PL3 broth, which contained large quantities of polysaccharides, significantly decreased the ratio of interferon-gamma (IFN- $\gamma$ ) to interleukin 4 (IL-4) in T cells. Thus, *P. linteus* fermented broths produced additive effects on the regulation of the Th1/Th2 balance and show promise for the development of immunomodulatory therapeutics.**

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[Key words: Cell cycle; Cytokine; *Phellinus linteus*; Polysaccharide; Th1/Th2 balance]

Traditional Chinese herbal medicines have been used for over a thousand years in East Asia, and have recently attracted much attention in Western countries. Mushrooms, as important ingredients in traditional Chinese medicines, have been shown to provide health benefits, including anti-inflammatory and immunomodulatory activities (1,2). *Phellinus linteus*, a species of mushroom in the family *Hymenochaetaceae* in the class *Basidiomycetes*, is indigenous to tropical South America, Africa, and East Asia, has attracted increasing attention in the last decade (2).

*P. linteus* has been used for its medicinal effects in the treatment of allergies, diabetes, gastroenteric dysfunction, and hemorrhage (2,3), and it shows the potential for development of anticancer therapy (4,5). In addition to fruiting body extracts of *P. linteus*, polysaccharides have shown to possess anti-oxidative and anti-inflammatory properties (6). Moreover, polysaccharides isolated from *P. linteus* have been demonstrated to inhibit the production of inflammatory cytokines in T helper 1 (Th1) cells and thus to attenuate the progression of autoimmune diabetes in non-obese

diabetic mice (7). These findings indicate that *P. linteus* has multiple applications in medicinal treatments.

Dysregulation of cytokine production by Th1 versus Th2 cells and the pro-inflammatory/anti-inflammatory balance may lead to human diseases (8). Our recent study demonstrated that *P. linteus* fermented broths modulated innate immunity by suppressing inflammatory responses in macrophages (9). However, no detailed study has been published to date on the regulation of T cells by *P. linteus* fermented broths, or exploring their relevance for clinical applications. In this study, several *P. linteus* fermented broths were prepared and their biological activity for the regulation of cell cycle progression in T cells was evaluated. Additionally, their regulatory effect on the cytokine production of T cells and the Th1/Th2 balance were investigated.

### MATERIALS AND METHODS

***P. linteus* mycelial materials, preparation, and purification** *P. linteus* mycelial fermented broths were prepared by Yushen Biotechnology Co., Ltd (Taichung, Taiwan). Briefly, four different compositions of broths (PL1–4) were prepared to culture *P. linteus* as described by our previous study (9). The mycelia were incubated in broth (pH5.2) at 28°C for 10 days with 125 rpm continuous shaking. The cultured broths were prepared by filtrations. The dialyzate medium (MD) which was prepared from PL1 culture broth and dialyzed against de-ionized water at 4°C for 72 h by using dialysis tubing with a molecular weight cutoff of 12,000–14,000. The broth filtrates and dialyzate were lyophilized for experimental use.

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**TABLE 1.** Contents of polysaccharides in *Phellinus linteus* fermented broths.

Tested sample <sup>a</sup>	Polysaccharides (mg/L)
PL1	103
PL2	82
PL3	172
PL4	115
MD	98

<sup>a</sup> The preparation of *P. linteus* fermented broths (PL1-4) and dialysate medium (MD) were described in Materials and methods.

**Determination of polysaccharides in culture broths** To determine the concentrations of polysaccharides in the samples, the broth filtrates and dialyzate were precipitated in four volumes of 95% (v/v) ethanol and incubated at 4°C for 24 h, then centrifuge at 13,000 ×g for 15 min. The pellet was suspended in 1 M NaOH at 60°C for 1 h, and total polysaccharide was then determined by phenol–sulfuric acid method (10).

**Cell culture** Murine lymphoma cell line, EL-4 cells (TIB-39) were purchased from American Type Culture Collection (ATCC, Rockville, MD, USA). Cells were cultured in RPMI 1640 medium supplemented with 10% fetal bovine serum (Hyclone Laboratories, Logan, UT, USA), and incubated at 37°C in 5% CO<sub>2</sub> humidified air atmosphere.

**Cytotoxicity assay** The mitochondrial respiration-dependent MTT assay was used to determine the cytotoxicity of *P. linteus* fermented extracts on EL-4 cells. MTT (0.1 mg in PBS) was added into each well and incubated at 37°C for 3 h. The MTT formazan crystals [1-(4,5-dimethylthiazol-2-yl)-3,5-diphenylformazan] were dissolved in 100 µl DMSO, and the mean OD of each well was used for assessing the cell viability and expressed as percentage of control.

**Analysis of cell cycle** EL-4 cells were cultured with the assigned concentrations of *P. linteus* fermented extracts for 24 h. The cells were washed with PBS and fixed in 75% alcohol at –20°C for 2 h. The prepared cells were stained with 20 µg/µl propidium iodide containing 0.1% Triton X-100 and 0.1 mM EDTA for 30 min at 4°C, and then analyzed by a FACS Calibur cytometer (Becton–Dickinson, San Jose, CA, USA) as described previously (11). The data were analyzed using Cell Quest software WinMDI (Verity Software House, Topsham, ME, USA).

**Determination of cytokine secretions** EL-4 cells were cultured with 10 ng/ml phorbol 12-myristate 13-acetate (PMA) or various concentrations of *P. linteus* fermented extracts for 24 h. The culture media were centrifuged at 13,000 ×g for 10 min, and culture supernatant were collected for further analysis. The levels of IL-2, IL-4, and INF-γ in supernatants were then analyzed using a sandwich enzyme-linked immunosorbent assay (ELISA) kit (R&D Systems, Minneapolis, MN, USA), according to the manufacturer's instruction.

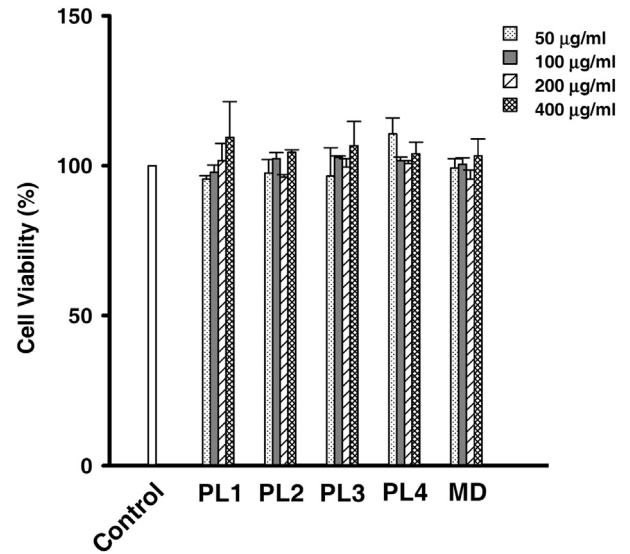
**Reverse transcription and real-time quantitative PCR** Total RNA from EL-4 cells was isolated by using TRIzol (Invitrogen, Carlsbad, CA, USA), as described previously with a slight modification (12). Briefly, total RNA (1 µg) was reverse-transcribed into cDNA by using oligo(dT) primers and Moloney Murine Leukemia Virus reverse transcriptase (Invitrogen). The RNA expression levels of T-bet and GATA-3 were determined by reverse transcription quantitative PCR (RT-qPCR), using StepOnePlus Real-Time PCR Systems (Applied Biosystems, Foster City, CA, USA). The oligonucleotide primers used corresponded to T-bet (forward: 5'-TGCCAGGAAACCGCTTATAG-3'; and reverse: 5'-AACTTCCTGGCCATCCA-3'), GATA-3 (forward: 5'-GGTGGACGTACTTTTAAACATCGA-3'; and reverse: 5'-CGTAGCCCTGACGGAGTTTC-3'), and glyceraldehyde-3-phosphate dehydrogenase (GAPDH: forward, 5'-ATGTGTCCGTCGTGGATCTGA-3'; and reverse: 5'-CCTGTTCCACCACCTTCTTGA-3') were synthesized by Invitrogen. After pre-incubation at 95°C for 20 s, PCR was performed with 40 cycles of 95°C for 18 s and 60°C for 30 s. The threshold was set above the non-template control background and within the linear phase of target gene amplification in order to calculate the cycle number at which the transcript was detected (denoted as [C<sub>T</sub>]).

**Statistical analysis** All data are presented as mean ± standard deviation of triplicate experiments. Statistical significance analysis was performed using Student's *t*-test; a *P* value <0.05 was considered significant.

## RESULTS

**Suppression of mitogen-induced cell cycle progression in T cells by *P. linteus*** *P. linteus* fermented broths were prepared and characterized as described in our previous study (9). The polysaccharide contents of each *P. linteus* fermented broth were analyzed. The PL1, PL2, PL3, PL4, and MD samples contained 82 mg/L, 103 mg/L, 172 mg/L, 115 mg/L, and 98 mg/L of polysaccharides, respectively (Table 1).

We then analyzed whether *P. linteus* fermented broths have cytotoxic effects on T cells. Cell viability testing was performed after



**FIG. 1.** *Phellinus linteus* fermented broths did not influence the proliferation of T cells. EL-4 cells were incubated with *P. linteus* fermented broths at the indicated concentrations (50, 100, 200, 400 µg/ml) for 24 h. The culture supernatants were collected and the effect of each tested sample on the proliferation of EL-4 cells was assessed by MTT assay. The data showed that there was no loss of cell viability in EL-4 cells during the incubation period. The results are shown as means ± standard deviations for three independent experiments.

EL-4 cells were incubated with various concentrations (50–400 µg/ml) of *P. linteus* fermented broths for 24 h. As shown in Fig. 1, the *P. linteus* fermented broths showed no cytotoxic effects on the cells. Therefore, a maximal concentration (400 µg/ml) of *P. linteus* fermented broth was used to perform the following experiments.

To analyze whether the growth inhibition of T cells in response to *P. linteus* fermented broths was associated with inhibition of cell activation, cell proliferation and cell cycle progression were analyzed in EL-4 cells following treatment with phorbol 12-myristate 13-acetate (PMA), a potent mitogen that activates T cells (13). As shown in Supplementary Fig. S1, following treatment of cells with PMA, cell proliferation increased. However, in cells treated with PL2, PL3, PL4, and MD, PMA-induced cell proliferation was significantly inhibited. We then investigated whether *P. linteus* fermented broths affect cell cycle progression. In the absence of treatment with PMA, a large proportion of the cells were found to be in G1 phase (Table 2). With PMA treatment, the proportion of cells in G1 phase was significantly suppressed. Noticeably, PMA-induced cell cycle progression was inhibited by PL3 and MD, mostly through arrest of cells in the S phase.

### Inhibitory effects of *P. linteus* on mitogen-induced IL-2 production in T cells

To determine whether the modulation of cell cycle progression in T cells by *P. linteus* was associated with cytokine stimulation, the production of IL-2 was measured. As shown in Fig. 2, PMA enhanced IL-2 production in T cells to a

**TABLE 2.** Attenuation of cell cycle progression in T cells by *Phellinus linteus* fermented broths.

Tested sample	G1	S	G2/M
Control	58.36 ± 1.5	16.18 ± 2.6	24.19 ± 1.6
PMA	43.74 ± 2.9	20.71 ± 1.1	31.30 ± 0.6
PL1	45.49 ± 1.0	19.98 ± 0.5	31.45 ± 0.7
PL2	45.16 ± 1.9	21.98 ± 1.1	30.06 ± 1.5
PL3	45.55 ± 0.1	25.00 ± 0.4 <sup>a</sup>	27.37 ± 1.8 <sup>a</sup>
PL4	59.48 ± 0.8 <sup>a</sup>	17.41 ± 2.1	22.25 ± 1.1 <sup>a</sup>
MD	57.60 ± 1.6 <sup>a</sup>	23.28 ± 2.2 <sup>a</sup>	20.11 ± 3.9 <sup>a</sup>

<sup>a</sup> *P* < 0.05 compared with PMA treatment.

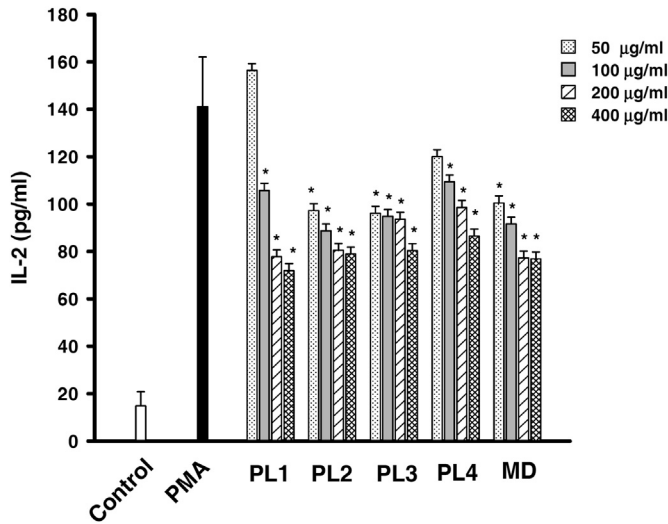


FIG. 2. Inhibition of mitogen-induced IL-2 production in T cells by *Phellinus linteus* fermented broths. EL-4 cells were incubated with PMA (10 ng/ml) followed by treatment with *P. linteus* fermented broths at the indicated concentrations (50, 100, 200, 400 µg/ml) for 24 h. The culture supernatants were prepared for analysis of IL-2 production by ELISA (R&D Systems) as described in Materials and methods. The results are shown as means ± standard deviations for three independent experiments. \**P* < 0.05 is considered significant.

high level, compared to the untreated control group. Treatment of PMA-stimulated EL-4 cells with *P. linteus* fermented broths significantly suppressed the enhanced IL-2 production in a concentration-dependent manner. At a low concentration of 50 µg/ml, PL2, PL3, and MD were the most potent inhibitors of IL-2 production. This result is concordant with the results of cell cycle analysis (Table 2), indicating that PL3 and MD can modulate T cell activation.

**Regulation of Th1/Th2 cytokine production in mitogen-activated T cells by *P. linteus*** We further investigated whether the *P. linteus* fermented broths affected the Th1/Th2 cytokine production of mitogen-activated T cells. Our results showed that PL3, PL4, and MD inhibited IFN-γ production in a concentration-dependent manner (Table 3). Moreover, at a concentration of 400 µg/ml, PL3 decreased the ratio of IFN-γ to IL-4 by 62% upon compared to PMA treatment alone. To confirm our findings, we

TABLE 3. Modulation of Th1/Th2 cytokine production by *Phellinus linteus* fermented broths.

Tested sample	IFN-γ (pg/ml)	Ratio <sup>a</sup>	IL-4 (pg/ml)	Ratio <sup>b</sup>	IFN-γ/IL-4 ratio
Mock	31.2 ± 5.7		14.9 ± 1.2		
PMA	462.9 ± 47.5	1.0	190.3 ± 16.8	1.0	1.00
PL1					
400	453.9 ± 17.4	1.0	191.6 ± 14.0	1.0	0.97
200	566.0 ± 22.1	1.2	181.4 ± 14.8	1.0	1.28
PL2					
400	441.1 ± 23.1	1.0	331.5 ± 28.1	1.7	0.55 <sup>c</sup>
200	521.7 ± 75.7	1.1	303.7 ± 20.3	1.6	0.71 <sup>c</sup>
PL3					
400	436.4 ± 21.8	0.9	477.5 ± 29.4	2.5	0.38 <sup>c</sup>
200	459.4 ± 41.8	1.0	239.1 ± 19.8	1.3	0.79 <sup>c</sup>
PL4					
400	394.2 ± 21.5	0.9	198.0 ± 15.2	1.0	0.82 <sup>c</sup>
200	432.0 ± 59.3	0.9	181.5 ± 15.9	1.0	0.98
MD					
400	352.9 ± 47.4	0.8	193.1 ± 16.7	1.0	0.75 <sup>c</sup>
200	382.5 ± 23.4	0.8	174.8 ± 13.6	0.9	0.90

<sup>a</sup> Ratio relative to PMA treatment.  
<sup>b</sup> Ratio relative to PMA treatment.  
<sup>c</sup> *P* < 0.05 compared with PMA treatment.

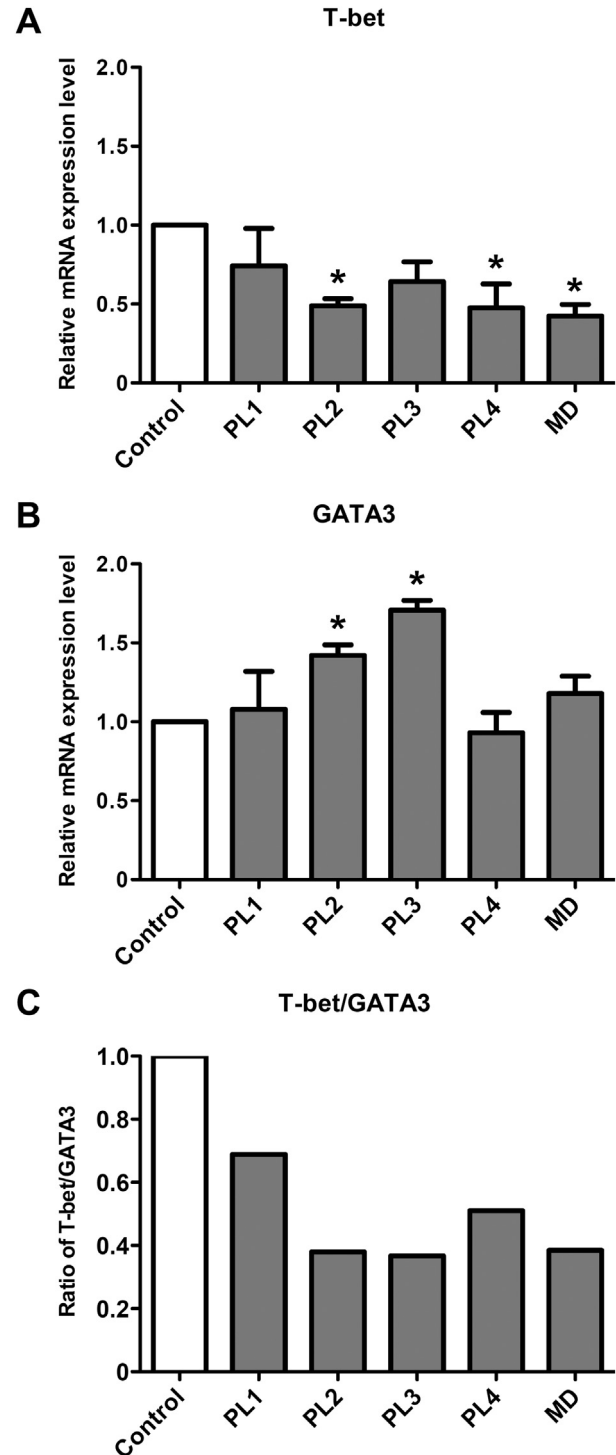


FIG. 3. Effects of *Phellinus linteus* fermented broths on the expression of T-bet and GATA-3. EL-4 cells were incubated with PMA (10 ng/ml) and *P. linteus* fermented broths (400 µg/ml) for 24 h. Total RNA isolated from EL-4 cells was prepared and the mRNA expression levels of (A) T-bet and (B) GATA-3 were detected by reverse transcription quantitative real-time PCR. (C) The ratio of T-bet/GATA-3 was determined. The results are shown as means ± standard deviations for three independent experiments. \* *P* < 0.05 compared to control group, as determined by Student's *t*-test.

then analyzed the mRNA expression levels of T-bet and GATA-3 in EL-4 cells. As shown in Fig. 3, treatment with PL2 and PL3 significantly increased the mRNA level of GATA-3. In particular, PL3 reduced the T-bet/GATA-3 ratio, which was significantly lower in the PL3 group than in the other treatment groups

(Fig. 3C). Collectively, these results indicate that *P. linteus* fermented broths, particularly PL3, produce an additive effect on the regulation of the Th1/Th2 balance through modulation of the ratio of IFN- $\gamma$  to IL-4.

## DISCUSSION

In this study, we found that five samples (PL1-4 and MD) derived from *P. linteus* fermented broths showed different immunoregulatory effects. As shown in Fig. 2, all tested samples showed comparable activity in inhibition of PMA-stimulated IL-2 production. We further found that PL3, MD, and PL4 significantly attenuated cell cycle progression in T cells (Table 2). PL3 and MD arrested the cell cycle at both the S and G2/M phases, whereas PL4 arrested it at the G1 phase. The effects of *P. linteus* fermented broths on Th1/Th2 regulation were then analyzed using ELISA. Our results showed that PL3, PL2, MD, PL4, and PL1 decreased the ratio of IFN- $\gamma$ /IL-4 by 62%, 45%, 25%, 18%, and 3%, respectively (Table 3). Additionally, PL3 showed the highest activity for reduction of the T-bet/GATA-3 ratio (Fig. 3). Notably, PL3 contained large quantities of polysaccharides (Table 1), which have been shown to strongly suppress mitogen-induced T cell activation. Several products isolated from *P. linteus*, including polysaccharides, proteoglycans, and glycoproteins, have been extensively studied to determine their immunomodulatory properties (6,14,15). Polysaccharides have been shown to possess the ability to modulate T cell immune responses (16–18). Our results are in accord with the findings of previous studies, and indicate that the polysaccharide contents of PL3 may contribute to its immunoregulatory function.

Oral administration of proteoglycans derived from *P. linteus* prevents collagen-induced arthritis in mice, by reducing production of antibodies and cytokines (19). Additionally, acidic polysaccharides isolated from *P. linteus* alleviate septic shock, associated with decreased expression of MHC II in B cells and macrophages (17). Our recent study demonstrated that *P. linteus* fermented broths act as potent anti-inflammatory agents by attenuating macrophage activation (9). In the present study, we showed that *P. linteus* fermented broths can inhibit mitogen-induced cell cycle progression and IL-2 production in T cells. Moreover, cytokine production by T cells and the Th1/Th2 balance can be regulated by *P. linteus* fermented broths. These data indicate that *P. linteus* possesses various biological activities, not only in regulation of innate immunity but also in modulation of acquired immunity, including humoral and cell-mediated responses, and that it shows potential for future development of therapeutic agents.

Dysregulation of the Th1/Th2 balance may be associated with several human diseases, including autoimmune disorders and allergic reactions (8). Moreover, uncontrolled Th1 responses may induce tissue damage and lead to increased disease severity (20), indicating that suppression of Th1 activation may be beneficial in attenuation of disease progression. For instance, inhibition of IL-12 activity reduced pathogenic Th1 responses and induced remission of human diseases including Crohn's disease, endotoxin-induced shock, and multiple sclerosis (21). In an *in vivo* study, treatment of non-obese diabetic (NOD) mice with polysaccharide isolated from *P. linteus* inhibited the expression of IFN- $\gamma$  and IL-2 by Th1 cells, but up-regulated IL-4 expression by Th2 cells, successfully attenuating the development of autoimmune diabetes (7). Additionally, proteoglycan from *P. linteus* showed the ability to alter the Th1/Th2 ratio, resulting in the prevention of autoimmune joint inflammation in mice (19). Our results are concordant with previous reports that *P. linteus* fermented broths regulate the Th1/Th2 balance and might be beneficial for the development of immunomodulation therapeutics.

In conclusion, our data demonstrate that *P. linteus* fermented broth exhibits immunomodulatory activity. *P. linteus* fermented

broth PL3 contained the highest quantity of polysaccharides and showed the strongest effects, indicating the immunomodulatory activity of this component. Although the detailed mechanisms underlying this effect require further investigation, this study suggests that *P. linteus* fermented broth is a promising remedy for immunomodulation through regulation of the Th1/Th2 balance.

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.jbiosc.2015.05.008>.

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